

Human hyaluronan ELISA Kit

(Catalog Number:31B110)

For the quantitative determination of human hyaluronan concentrations in serum, plasma or cell culture supernate samples

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INTRODUCTION

Hyaluronan, also known as hyaluronic acid (HA) or sodium hyaluronate, is a naturally occurring linear polymer. Hyaluronan is a glycosaminoglycan (GAG) that is ubiquitously present in the extracellular matrix of all vertebrates and is also present in the capsule of some strains of Streptococci. Mammalian hyaluronan is synthesized by one of three distinct hyaluronan synthases (HAS1, 2, and 3), which produce HA polymers with different chain lengths and differ in their rates of synthesis. Whereas high molecular weight HA (> 500 kDa) is anti-angiogenic, anti-inflammatory, and immunosuppressive, low molecular weight HA (10-500 kDa) is highly angiogenic and pro-inflammatory.[1] Functionally, hyaluronan molecules are important for the maintenance of a highly hydrated extracellular matrix in tissues, which is involved in cell adhesion and tissue fibrogenesis. Thus, hyaluronan is a potential biomarker for fibrosis detection.

PRINCIPLE OF THE ASSAY

This assay is a quantitative sandwich enzyme-linked immunosorbent assay (ELISA). The microtiter plate is pre-coated with a polyclonal antibody specific for human hyaluronan. Standards and samples are pipetted into the wells and any human hyaluronan present is bound by the immobilized antibody. After washing away any unbound substances, a biotin labelled polyclonal antibody specific for human hyaluronan is added to the wells. After wash step to remove any unbound reagents, streptavidin-horseradish peroxidase conjugate (STP-HRP) is added. After the last wash step, an HRP substrate solution is added and color develops in proportion to the amount of human hyaluronan bound initially. The assay is stopped, and the optical density of the wells is determined using a microplate reader. Since the increases in absorbance are directly proportional to the amount of captured human hyaluronan, the unknown sample concentration can be interpolated from a reference curve included in each assay.

INTENDED USE

This Human hyaluronan ELISA kit is designed for quantification of human hyaluronan in serum, plasma, and cell culture supernate samples.

REAGENTS SUPPLIED

Each kit is sufficient for one 96-well plate and contains the following components:

1. Microtiter Strips (96 wells), coated with a polyclonal antibody against human hyaluronan, sealed
2. 10×Wash buffer, 50 mL
3. 5×Assay buffer, 20 mL
4. 100×Detection antibody solution, a biotin-labeled polyclonal antibody against human hyaluronan, 0.12 mL
5. Human hyaluronan standard, 90 ng of human hyaluronan, lyophilized
6. 200×STP-HRP solution, 0.06 mL

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7. Substrate solution, 12 mL, ready for use
8. Stop solution, 12 mL, ready for use

OTHER MATERIALS REQUIRED, BUT NOT PROVIDED

1. Pipettes and pipette tips
2. 96-well plate or manual strip washer
3. Buffer and reagent reservoirs
4. Paper towels or absorbent paper
5. Plate reader capable of reading absorbency at 450 nm
6. Distilled water or deionized water

STORAGE

The kit should be stored at 2-8°C upon receipt, and all reagents should be equilibrated to room temperature before use. Remove any unused antibody-coated strips from the human hyaluronan microtiter plate, return them to the foil pouch and re-seal. Once opened, the strips may be stored at 2-8°C for up to one month.

PREPARATION OF REAGENTS

Bring all reagents and materials to room temperature before assay.

A. 1×Assay buffer

Prepare 1×Assay buffer by mixing the 5×Assay buffer (20 mL) with 80 mL of distilled water or deionized water. If precipitates are observed in the 5×Assay buffer bottle, warm the bottle in a 37°C water bath until the precipitates disappear. The 1×Assay buffer may be stored at 2-8°C for up to one month.

B. 1×Wash buffer

Prepare 1×Wash buffer by mixing the 10×Wash buffer (50 mL) with 450 mL of distilled water or deionized water. If precipitates are observed in the 10×Wash buffer bottle, warm the bottle in a 37°C water bath until the precipitates disappear. The 1×Wash buffer may be stored at 2-8°C for up to one month.

C. 1×Detection antibody solution

Spin down the 100×Detection antibody solution briefly and dilute the desired amount of the antibody 1:100 with 1×Assay buffer, 100 µL of the 1×Detection antibody solution is required per well. Prepare only as much 1×Detection antibody solution as needed. Return the 100×Detection antibody solution to 2-8°C immediately after the necessary volume is removed.

D. 1×STP-HRP solution

Spin down the 200×STP-HRP solution briefly and dilute the desired amount of the 200×STP-HRP solution 1:200 with 1×Assay buffer, 100 µL of the 1×STP-HRP solution is required per well. Prepare only as much 1×STP-HRP solution as needed. Return the 200×STP-HRP solution to 2-8°C immediately after the necessary volume is removed.

PREPARATION OF STANDARDS AND SAMPLES

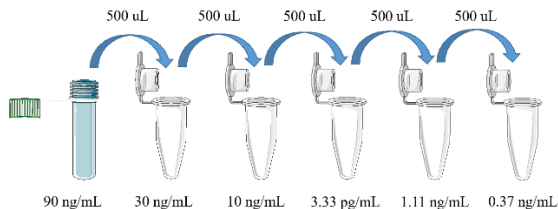
Human hyaluronan Standards: Reconstitute the lyophilized standard with 1.5 mL of 1×Assay buffer to generate a standard stock solution of 90 ng/mL. Allow the standard

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to sit for 10 minutes with gentle agitation prior to making dilutions. Pipette 1000 μL of 1 \times Assay buffer to 30, 10, 3.33, 1.11, 0.37ng/mL tubes. Use the standard stock solution to produce a serial dilution as shown below.



1 \times Assay buffer serves as the zero standard (0 pg/mL). The reconstituted standard stock should be aliquoted and stored at -80°C for up to one month. Avoid repeating freezing/thawing cycles. Please do not store the diluted standard solutions.

Sample Preparation:

Serum or plasma sample generally requires a **5-fold** dilution in the 1 \times Assay buffer. It is recommended that the users establish their own dilution factors based on the concentration range of their samples.

ASSAY PROCEDURE

It is recommended that all standards and samples be assayed in duplicate.

1. Add 100 μL of standard or sample per well, and incubate at room temperature for 2 hours.
2. Discard the content and tap the plate on a clean paper towel to remove residual solution in each well. Add 300 μL of 1 \times Wash buffer to each well and incubate for 1 minute. Discard the 1 \times Wash buffer and tap the plate on a clean paper towel to remove residual wash buffer. Repeat the wash step for a total of 3 washes.
3. Add 100 μL of 1 \times Detection antibody solution to each well, and incubate at room temperature for 2 hours.
4. Wash each well 3 times as in step 2.
5. Add 100 μL of 1 \times STP-HRP solution to each well, and incubate at room temperature for 30 minutes.
6. Wash each well 4 times as described in step 2.
7. Add 100 μL of Substrate solution to each well, and incubate at room temperature for 15 minutes. **Protect from light.**
8. Add 50 μL of Stop solution to each well, and gently tap the plate frame for a few seconds to ensure thorough mixing.
9. Measure the absorbance of each well at 450 nm immediately.

CALCULATION

1. Subtract the absorbance of the blank from that of standards and samples.
2. Generate a standard curve by plotting the absorbance obtained (y-axis) against human hyaluronan concentrations (x-axis). The best-fit line can be generated with

any curve-fitting software by regression analysis. Any curve of 4-parameter or log-log curve fitting can be used for calculation.

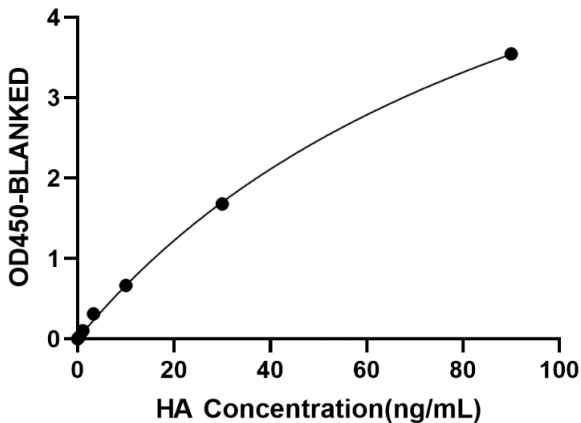
- Determine the human hyaluronan concentration of samples from the standard curve and multiply the value by the dilution factor.

TYPICAL STANDARD CURVE

The following standard curve is provided for demonstration only. A standard curve should be generated for each set of sample assays.

Human hyaluronan (ng/mL)	Absorbance (450 nm)	Blanked Absorbance
90.00	3.622	3.545
30.00	1.755	1.678
10.00	0.742	0.665
3.33	0.387	0.31
1.11	0.177	0.1
0.37	0.102	0.025
0	0.077	0

Human hyaluronan standard curve (4-parameter)



ASSAY CHARACTERISTICS

A. Sensitivity

The lowest level of human hyaluronan that can be detected by this assay is 0.12ng/mL.

B. Precision

Intra-assay Precision (Precision within an assay) C.V. <3.6%.

Inter-assay Precision (Precision between assays) C.V. <5.8%.

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2. Ralf Lichtinghagen, et al. (2012) *The Enhanced Liver Fibrosis (ELF) score: Normal values, influence factors and proposed cut-off values.*

SUMMARY OF ASSAY PROCEDURE

